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Total Synthesis of α-Deamino-3-(β-Dglucopyranosyloxy)kynurenine

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Abstract— α -Deamino-3-(β -D-glucopyranosyloxy)kynurenine, a yellow, fluorescent compound isolated from human lens, was synthesized in 8 steps (10% overall yield) from commercially available 3-methoxy-2-nitrobenzaldehyde. Key events included preparation and glucosylation of methyl 4-(3-hydroxy-2-nitrophenyl)-4-oxobutanoate followed by reduction of the nitro group and deprotection. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

 α -Deamino-3-(β -D-glucopyranosyloxy)kynurenine (1) is the most recently identified member of a series of UV-filtering agents isolated from the protein-free extract of human eye lens.¹ Other known members of this class of compound are 3-(β -D-glucopyranosyloxy)-L-kynurenine (**2**),^{2,3} kynurenine (**3**),^{2,3} and 3-hydroxy-L-kynurenine (**4**).⁴ A structurally related glucoside, 2-amino-3-(β -D-glucopyranosyloxy)acetophenone (5),⁵ has been isolated from the insoluble protein fraction of human lens. The three glucosides are unusual both because glucoside formation in mammalian tissue is rare and because these compounds are found in the lenses of primates but not in the lenses of other animals.² Although compounds 1-5 may serve as photoprotective filters,⁶ they limit the frequency range of visual perception by primates⁷ and may contribute to cataract formation through photosensitization of molecular oxygen.^{2,8} Access to these compounds in order to study their biological and photophysical properties is clearly desirable.



Keywords: glycosidation; lithiation; natural products; NMR.



To provide material in a quantity suitable for further study, we have devised a practical synthesis of 1. During the course of our work, another synthesis of 1 was also reported.⁹

Results and Discussion

We initially investigated *ortho*-metalation/acylation of a suitably protected 2-aminophenol, such as 7 (Scheme 1), as a route to assemble the aglycone portion of **1**. Although the dianion of **7** could be generated (3.0 equiv. of *t*BuLi in ether, -78 to -20° C) and successfully formylated or deuterated, reactions of the dilithium salt of **7** with active acyl derivatives (anhydride, acid chloride, and Weinreb amide¹⁰) of succinic acid led only to recovered **7** (Table 1). Apparently, the hard aryl anion prefers to deprotonate the acyl



Scheme 1.

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Table 1. ortho-Metalation reactions of 7





donor. Treatment of the dilithium salt of 7 with 1 equiv. of succinyl chloride prevented deuterium incorporation in the aromatic ring upon subsequent quench and work-up with D_2O (Table 1, entries 2 and 5). This behavior stands in stark contrast to the usual tendency of organolithium reagents to add efficiently not only to acid chlorides but also to the ketones thus produced.¹⁰⁻¹² Also surprising is the apparent failure of succinyl chloride to undergo dehydrohalogenation and generate a ketene intermediate, which itself would be a highly reactive acylating agent.¹³ Although a softer aryl anion, such as a magnesium, manganese, zinc, or copper salt, would probably undergo succinylation satisfactorily,¹⁴ we also found that protected aminophenol 10 was not readily glycosylated by tetra-O-acetylglucopyranosyl bromide (9) in the presence of silver imidazolate¹⁵ (Scheme 2). We therefore turned to an aglycone intermediate in which a nitro group served as the precursor to the amino group.

In a model study, glucosylation of 2-nitrophenol (12) proceeded smoothly to give aryl glucoside 13 in 76%

yield. Apparently, this glycosylation requires the phenoxide anion as the nucleophile, and silver imidazolate is able to deprotonate **12** but not **10**.

Toward synthesizing the actual aglycone, we set about to allylate 3-methoxy-2-nitrobenzaldehyde (14). Although allylmagnesium bromide failed to give the desired product in reasonable yield, reaction of 14 with allyltrimethylsilane¹⁶ in the presence of TiCl₄ gave 15 in 90% yield (Scheme 3). While hydroboration of 15 as the free alcohol¹⁷ using BH₃·THF or 2 mol equivalents of dicyclohexylborane generated in situ¹⁸ failed to generate 16 satisfactorily,¹⁹ exposure of 15 to a slight excess of borane (previously treated with a substoichiometric amount of cyclohexene) followed by basic H₂O₂ produced diol 16 in good yield and high regioselectivity. Oxidation of 16 with Jones reagent gave the desired ketoacid in modest yield. Demethylation of 17 with LiCl in DMF at reflux²⁰ followed by esterification of the reaction residue by BF₃·OEt₂ in methanol at reflux produced the desired aglycone 18.

Coupling of **18** and **9** proceeded efficiently in the presence of silver imidazolate and zinc chloride to afford glucoside **19** as a single diastereomer (Scheme 4). Catalytic reduction of **19** over Pd-C gave **20**, which was deacetylated in methanol that had not been previously dried to give the free acid, **1**, directly.

The identity of **1** was established by agreement of its NMR data with the partial ¹H and ¹³C spectra reported. ¹ The structure of **1** was further confirmed and its ¹H and ¹³C resonances were assigned by NOE difference, COSY, HMQC, and HMBC experiments.

Conclusion

In conclusion, we have developed a practical synthesis of fluorescent glucoside **1**, confirmed its structure, and made full assignment of its ¹H and ¹³C NMR spectra. Further study is required to determine the role of **1** and related glucosides as UV filters in human lens and as possible causative agents in the aging of human lens, particularly in the generation of cataracts.

Experimental

General procedures

Methylene chloride and THF were purified by distillation under nitrogen from CaH_2 or sodium/benzophenone, respectively. All reactions were conducted under nitrogen in flame or oven-dried glassware with magnetic stirring, unless noted





Scheme 3.

otherwise. TLC was performed on 2.5×10 cm plates coated with 250 µm-thick silica gel GF. Column chromatography was performed on flash-grade silica gel (60 Å, 230–400 mesh).

Unless specified otherwise, ¹H NMR and ¹³C NMR spectra (400 and 101 MHz, respectively) were recorded using CDCl₃ as the solvent. A JVERT pulse sequence differentiated ¹³C nuclei bearing an even number of hydrogen atoms from those bearing an odd number. IR spectra were obtained of samples as neat oils or KBr pellets. Liquid secondary ion mass spectra (LSIMS) were obtained using Cs⁺ (15 eV) as the ionizing beam and either glycerol or *p*-nitrobenzyl alcohol as the matrix. In some cases, NaI was added to the matrix to enhance the relative intensity of the quasi-molecular ion. Elemental analysis was performed by Micro-Analysis, Inc.

1-(3-Methoxy-2-nitrophenyl)-3-buten-1-ol (15). To a solution of 3-methoxy-2-nitrobenzaldehyde (3.62 g, 20 mmol) in 20 mL of CH_2Cl_2 was added $TiCl_4$ (10 mL of

a 1 M solution in CH₂Cl₂) over 15 min. After 10 min at rt, 4.8 mL of allyltrimethylsilane (3.45 g, 30 mmol) was added all at once. After 10 more min, the reaction mixture was diluted with ether, washed with water and brine, dried (Na₂SO₄), and concentrated. Bulb-to-bulb distillation of the residual brown oil (5 Torr, bath temperature=160°C) gave **15** as a pale yellow oil (4.10 g, 92%). $R_{\rm f}$ (1:2 EtOAc-hexanes) 0.65. ¹H NMR δ 2.19 (d, J=3.2 Hz, 1H), 2.47 (m, 1H), 2.59 (m, 1H), 3.89 (s, 3H), 4.73 (m, 1H), 5.15 (m, 2H), 5.78 (m, 1H), 6.96 (d, J=8.3 Hz, 1H), 7.17 (d, J=7.7 Hz, 1H), 7.44 (dd, J=7.7, 8.3 Hz, 1H). ¹³C NMR (even) δ 43.3, 115.0, 119.8, 137.2, 150.9; (odd) δ 56.9, 69.0, 111.9, 118.9, 131.6, 133.9. IR (neat) 3418, 1852, 1609, 1531, 1438 cm⁻¹. HRLSIMS calcd for C₁₁H₁₄NO₄ (M+H⁺) 224.0923, found 224.0910.

1-(3-Methoxy-2-nitrophenyl)-1,4-butanediol (16). Cyclohexene (1.28 g, 16 mmol) and 15 mL of THF were combined in a 250-mL round-bottomed flask. The flask was cooled in an ice-water bath and BH_3 ·THF (24 mL of a 1 M solution in THF) was added. After 10 min, **15** (4.55 g,



20 mmol) in 15 mL of THF was added. The ice bath was removed and the reaction was stirred for 30 min while warming to rt. With caution, 10 mL of 3.0 M NaOH followed by 3.42 mL of 30% aqueous H₂O₂ were added over 10 min. The reaction was stirred vigorously for 5 min, diluted with ether, and washed sequentially with saturated aqueous NH₄Cl, water, and brine. The organic layer was dried (Na₂SO₄) and concentrated. Chromatography of the residue gave 16 as a pale yellow solid (3.69 g, 75%), mp 103°C. $R_{\rm f}$ (1:3 EtOAc-hexanes) 0.15. ¹H NMR δ 1.69 (m, 2H), 1.86 (m, 2H), 3.67 (m, 2H), 3.89 (s, 3H), 4.70 (m, 1H), 6.95 (d, J=8.3 Hz, 1H), 7.20 (d, J=7.8 Hz, 1H), 7.43 (dd, J=7.8, 8.3 Hz, 1H). ¹³C NMR (even) δ 29.5, 36.2, 63.0, 138.2, 140.2, 150.8; (odd) δ 56.9, 69.9, 111.7, 118.9, 131.6. IR (KBr) 3320, 1527, 1372, 1280, 1058, 1019 cm^{-1} . HRLSIMS calcd for $C_{11}H_{16}NO_5$ (M+H⁺) 242.1028, found 242.1027. Anal. calcd for C₁₁H₁₅NO₅: C, 54.77; H, 6.27; N, 5.81. Found: C, 54.91; H, 6.42; N, 5.80.

4-(3-Methoxy-2-nitrophenyl)-4-oxobutanoic acid (17). A solution of 0.10 g of CrO₃ in 2 mL of 80% aqueous H₂SO₄ was added dropwise over 5 min to a solution of 16 (360 mg, 1.49 mmol) in 15 mL of acetone. After 30 min, 5 g of silica gel was added, and the mixture was evaporated to dryness. The residue was placed on a column of silica gel and eluted to give 17 (208 mg, 55%) as pale brown needles, mp 139°C. $R_{\rm f}$ (1:9 acetone/CH₂Cl₂) 0.30. ¹H NMR (250 MHz) δ 2.80 (t, J=6.5 Hz, 2H), 3.23 (t, J=6.5 Hz, 2H), 3.93 (s, 3H), 7.25 (d, J=8.2 Hz, 1H), 7.42 (d, J=7.9 Hz, 1H), 7.55 (dd, J=7.9, 8.2 Hz, 1H). ¹³C NMR (DMSO-d₆) (even) δ 28.4, 40.0, 131.8, 140.2, 152.4, 174.0, 197.9; (odd) δ 57.8, 118.8, 122.2, 132.8. IR (KBr) 1698, 1695, 750, 668 cm^{-1} . HRLSIMS calcd for $C_{11}H_{12}NO_6$ (M+H⁺) 254.0664, found 254.0653. Anal. calcd for C₁₁H₁₁NO₆: C, 52.18; H, 4.38; N, 5.53. Found: C, 52.51; H, 4.41; N 5.59.

4-(3-hydroxy-2-nitrophenyl)-4-oxobutanoate Methyl (18). A solution of LiCl (80 mg, 1.89 mmol) and 17 (130 mg, 0.51 mmol) in 10 mL of 95% aqueous DMF was heated at reflux for 4 h. The reaction mixture was diluted with ethyl acetate, washed sequentially with 5% aqueous HCl, water, and brine, dried (Na₂SO₄), and concentrated. The residue was dissolved in 20 mL of dry methanol and 0.5 mL of BF₃·OEt₂ was added. The reaction was heated at reflux for 1 h, and then concentrated. The residue was chromatographed to give 18 (59 mg, 45%) as a yellow oil. $R_{\rm f}$ (1:3 EtOAc-hexanes) 0.32. ¹H NMR δ 2.83 (t, J=6.6 Hz, 2H), 3.09 (t, J=6.6 Hz, 2H), 3.73 (s, 3H), 6.94 (dd, J=1.1, 7.3 Hz, 1H), 7.23 (dd, J=1.1, 8.5 Hz, 1H), 7.61 (dd, J=7.3, 8.5 Hz, 1H), 10.51 (br s, 1H) 13 C NMR (even) δ 28.4, 38.1, 115.0, 140.3, 155.6, 173.4, 200.7; (odd) & 52.4, 119.0, 121.6, 137.4. IR (KBr) 1734, 1699 cm⁻¹. HRMS (EI) calcd for C₁₁H₁₁NO₆ 253.0586, found 253.0577.

3-(4-Methoxy-4-oxobutanoyl)-2-nitrophenyl tetra-*O*acetyl- β -D-glucopyranoside (19). To a mixture of 18 (32 mg, 0.126 mmol) and 9 (156 mg, 0.380 mmol) dissolved in 6 mL of CH₂Cl₂ was added silver imidazolate (66 mg, 0.377 mmol) followed by 0.38 mL of 1.0 M ZnCl₂ in ether. After stirring 16 h at rt in the dark, the reaction mixture was filtered, and the filtrate was concentrated. Chromatography of the residue gave recovered 18 (12 mg) followed by **19** as a yellow foam (38 mg, 83% based on starting material consumed). $R_{\rm f}$ (1:1 EtOAc-hexanes) 0.28. ¹H NMR δ 2.05 (s, 3H), 2.10 (s, 3H), 2.14 (s, 3H), 2.17 (s, 3H), 2.67–2.84 (m, 2H), 3.12–3.33 (m, 2H), 3.69 (s, 3H), 3.79–3.94 (m, 1H), 4.24–4.29, (m, 2H), 4.99–5.35 (m, 4H), 7.52–7.58 (m, 2H), 7.62–7.67 (m, 1H) ¹³C NMR (even) δ 27.7, 34.9, 61.6, 131.3, 140.8, 148.2, 169.2, 169.2, 170.1, 170.4, 172.7, 195.8; (odd) δ 20.4, 20.5, 20.7, 21.0, 51.9, 67.9, 70.2, 72.0, 72.3, 100.5, 123.6, 123.8, 131.1 IR (KBr) 1743, 1699 cm⁻¹. HRLSIMS calcd for C₂₅H₂₉NO₁₅Na (M+Na⁺) 606.1462, found 606.1449.

3-(4-Methoxy-4-oxobutanoyl)-2-aminophenyl tetra-Oacetyl-β-D-glucopyranoside (20). A solution of 6 (32 mg, 0.055 mmol) in 5 mL of methanol was stirred with 8 mg of 5% Pd-C under an atmosphere of H₂ at rt for 3 h. The reaction mixture was filtered and concentrated. Chromatography of the residue (1:3 EtOAc-hexane with 1% Et₃N) afforded 7 (26 mg, 86%) as a pale yellow foam. R_f (1:1 EtOAchexanes) 0.48. ¹H NMR δ 2.04 (s, 3H), 2.05 (s, 3H), 2.08 (s, 3H), 2.20 (s, 3H), 2.71 (m, 2H), 3.30 (t, J=6.6 Hz, 2H), 3.71 (s, 3H), 3.86 (ddd, J=2.4, 5.1, 9.9 Hz, 1H), 4.18 (dd, J=2.4, 12.3 Hz, 1H), 4.31 (dd, J=5.1, 12.3 Hz, 1H), 4.96 (m, 1H), 5.16 (m, 1H), 5.29–5.34 (m, 2H), 6.51 (br s, 2H), 6.55 (dd, J=7.8, 8.2 Hz, 1H), 7.04 (dd, J=1.0, 7.8 Hz, 1H), 7.53 (dd, J=1.0, 8.2 Hz, 1H). ¹³C NMR (even) δ 28.8, 34.1, 61.8, 117.8, 142.2, 144.6, 169.4, 169.9, 170.1, 170.5, 173.6, 199.7; (odd) δ 20.5, 20.6, 20.7, 20.8, 51.8, 68.2, 71.1, 72.1, 72.2, 100.3, 113.9, 118.9, 125.5. IR (KBr) 1748, 1689, 1640 cm⁻¹ HRLSIMS calcd for C₂₅H₃₁NO₁₃Na (M+Na⁺) 576.1693, found 576.1711.

 α -Deamino-3-(β -D-glucopyranosyloxy)kynurenine (1). To a solution of 7 (26 mg, 0.047 mmol) in 5 mL of methanol was added 0.20 mL of 0.025 M NaOMe in methanol. After 14 h at rt, the reaction mixture was concentrated. The residue was chromatographed on reverse-phase silica gel (Analtech, C_{18} -bonded) with water to give 8 (14 mg, 80%) yield) as a yellow crystalline solid. $R_{\rm f}$ (1:5:94 AcOH-EtOH-EtOAc) 0.22. ¹H NMR (D₂O) δ 2.49 (t, J=6.8 Hz, 2H, H α), 3.23 (t, J=6.8 Hz, 2H, Hβ), 3.48 (m, 1H, H4'), 3.55 (m, 1H, H5'), 3.58 (m, 1H, H3'), 3.60 (m, 1H, H2'), 3.74 (dd, J=5.5, 12.5 Hz, 1H, H6'a), 3.89 (dd, J=2.0, 12.5 Hz, 1H, H6'b), 4.99 (d, J=7.1 Hz, 1H, H1[']), 6.75 (dd, J=7.9, 8.3 Hz, 1H, H5), 7.28 (dd, J=1.1, 7.9 Hz, 1H, H4), 7.67 (dd, J=1.1, 8.3 Hz, 1H, H6). ¹³C NMR (D₂O) δ 32.2 (Cα), 36.2 (Cβ), 60.8 (C6'), 69.7 (C4'), 73.2 (C3'), 75.8 (C2'), 76.5 (C5'), 101.9 (C1[']), 116.2 (C5), 119.6 (C1), 120.7 (C4), 126.4 (C6), 141.4 (C2), 145.2 (C3), 182.3 (CO₂H), 204.7 (C=O). HRLSIMS (M+Na⁺) calcd for $C_{16}H_{21}NO_9Na$ 394.1114, found 394.1118.

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